RUNTOGEN

FREEZING PROTOCOL

Always freeze down cells at a high concentration and at as low a passage number as possible. Ensure that the cells are at least 90% viable before freezing.

Always use proper aseptic technique and work in a laminar flow hood. Always wear personal protective equipment when working with liquid nitrogen.

1. Harvest log phase cells (with > 90% viability):

   For adherent cells, gently detach the cells from the culture vessel to collect cells into a centrifuge tube following the Subculturing Protocol (step 1 to step 6).

   For suspension cells, harvest all cells into a centrifuge tube.

2. Determine viable cell density and calculate the required volume of Cryopreservation Medium needed. We recommend freezing cells at 1.0 to 2.5 x 10^6 cells/ml.

3. Centrifuge the cell suspension at 1500 rpm for 3 minutes.

4. Aseptically, aspirate out the supernatant without disturbing the pellet.

5. Re-suspend the cell pellet in Cryopreservation Medium at the appropriate cell densit y.

6. Dispense the cell suspension into cryovials and freeze according to your laboratory s tandard (i.e. controlled rate freezing at approximately 1°C decrease per minute).

7. Transfer the frozen cells into liquid nitrogen storage (in the gas phase above the liqui d nitrogen) for long-term storage.

For laboratory research only. Not for clinical applications.
For technical questions, please email us at info@runtogen.com

Last revised November 13, 2023

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